



# Internet and Print Resources to Facilitate Pathology Analysis When Phenotyping Genetically Engineered Rodents

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## Abstract

Genetically engineered mice and rats are increasingly used as models for exploring disease progression and mechanisms. The full spectrum of anatomic, biochemical, and functional changes that develop in novel, genetically engineered mouse and rat lines must be cataloged before predictions regarding the significance of the mutation may be extrapolated to diseases in other vertebrate species, including humans. A growing list of reference materials, including books, journal articles, and websites, has been produced in the last 2 decades to assist researchers in phenotyping newly engineered rodent lines. This compilation provides an extensive register of materials related to the pathology component of rodent phenotypic analysis. In this article, the authors annotate the resources they use most often, to allow for quick determination of their relevance to research projects.

## Keywords

functional genomics, Internet, mouse, phenogenomics, phenotype, rat, reference, website

In the past 25 years, genetically engineered mice (GEM) and rats (GER) have advanced from novelty status to essential tools in biomedical research programs.<sup>167,168,220</sup> The driving force behind this evolution is the conviction that the functional and structural consequences of altering a rodent gene will be mirrored in human patients who carry a similar genetic anomaly in the homolog gene.<sup>204</sup> Supporting the accuracy of this premise is the fact that the effects in human patients of the 100 top-selling pharmaceutical agents are predicted by the GEM phenotypes that arise when genes encoding each drug's target are deliberately disrupted.<sup>245</sup> Another development in genetic engineering during the last decade has been the generation of humanized rodent models (ie, substitution of a human molecule or cell type in place of its native rodent counterpart), thus providing a system to test the functional and structural significance of human molecules *in vivo*.<sup>168,243</sup> The push to annotate the whole mammalian genome by comprehensive gene targeting in GEM<sup>30,130,193</sup> will greatly expand the magnitude of engineered rodent research and its application to human disease modeling.

Proficient phenotyping of GEM and GER requires regular access to a range of biomedical expertise. The complexity of phenotypic analysis typically necessitates a team approach for greatest success. In many cases, such know-how is provided within an institution through the assembly of "comprehensive centers" or "shared phenotyping resources" comprising several collaborating research and/or service laboratories. In other instances, the entire institute is fully dedicated to rodent phenotyping. Either strategy is suitable as long as a critical mass of

multidisciplinary expertise is available to support the efforts of the research team conducting its experiments.

The key bottleneck affecting all rodent phenogenomic programs, including the international effort to annotate the mammalian genome,<sup>130</sup> is the inability to perform high-throughput phenotypic analysis at a pace that matches the generation of new GEM and GER lines. A chief factor in this phenotyping backlog is the well-documented dearth of experienced comparative pathologists.<sup>13,35</sup> This shortage is the outcome of 2 primary trends in scientific research. First, laboratory animal pathologists are usually allocated to other research roles, such as disease diagnosis in laboratory animal facilities or evaluation of rodent tissues from toxicity bioassays. More important, training programs designed to produce comparative pathologists with substantial experience in rodent-based research are uncommon, so most individuals who enter the field do so at present by undertaking an informal course of mentored or self-directed independent study.<sup>24</sup> The major flaw of this individualized approach to comparative pathology training is that,

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**Table 1.** Essential Printed Resources for Comparative Pathologists Engaged in Phenotypic Analysis of Engineered Rodents

Title	First Author/Editor	Type	Citation
<p>Analytic methods</p> <p>"Evaluating Mutant Mice: Anatomic Pathology"</p> <p>This brief review provides a succinct introduction to major considerations required to phenotype rodents. Emphasis is placed on basic knowledge and skills required for structural analysis of adult animals.</p>	Brayton	Article	27
<p><i>Mouse Phenotypes: A Handbook of Mutation Analysis</i></p> <p>This book is a good introduction regarding procedures used to generate and assess prenatal and postnatal phenotypes of mice.</p>	Papaioannou	Book	171
<p><i>Systematic Approach to Evaluation of Mouse Mutations</i></p> <p>This volume describes the basic characteristics of building and maintaining mouse colonies, as well as techniques used to evaluate mutation-induced phenotypes.</p>	Sundberg	Book	216
<p>Anatomy</p> <p><i>Anatomy of the Rat</i></p> <p>This guide provides a detailed, well-illustrated description of rat organs and systems. Most of the information can be applied to mice.</p>	Greene	Book	90
<p><i>Colour Atlas of Anatomy of Small Laboratory Animals</i></p> <p>This 2-volume set includes detailed color plates showing major anatomic features for many rodent species, including mice and rats.</p>	Popesko	Book <sup>a</sup>	187
<p>Biology of rodents</p> <p><i>The Laboratory Mouse</i></p> <p>This book reviews major aspects of mouse-based research, including history, husbandry, strain nomenclature, breeding, anatomy, physiology, pathology, pathogens, genomics, procedures, and safety study design.</p>	Hedrich	Book	96
<p><i>The Laboratory Rat</i></p> <p>This reference reviews most aspects of rat-based biomedical research, including history, husbandry, strain nomenclature, breeding, anatomy, physiology, procedures, and design of various safety studies.</p>	Krinke	Book	135
<p><i>The Laboratory Rat</i></p> <p>One of the "Blue Book" volumes commissioned by the American College of Laboratory Animal Medicine, this volume provides a detailed overview of rat biology, including multiple chapters on various disease conditions.</p>	Suckow	Book	213
<p><i>The Mouse in Biomedical Research</i>, 2nd ed.</p> <p>Vol 1: History, Wild Mice, and Genetics</p> <p>Vol 2: Diseases</p> <p>Vol 3: Normative Biology, Husbandry, and Models</p> <p>Vol 4: Immunology</p> <p>These 4 American College of Laboratory Animal Medicine "Blue Books" provide encyclopedic coverage of mouse biology and pathology, as well as the place of mouse models of disease in biomedical research.</p>	Fox	Books	70 71 72 73
<p>Clinical pathology</p> <p><i>The Clinical Chemistry of Laboratory Animals</i></p> <p>This resource offers an extensive review of principal clinical chemistry analyses and their interpretation from multiple laboratory animal species, including mice and rats.</p>	Loeb	Book	140

(continued)

**Table 1.** (continued)

Title	First Author/Editor	Type	Citation
<p><i>Schalhm's Veterinary Hematology</i>                      The volume provides a thorough overview of blood collection and analytic methods for many species, including detailed chapters on mouse and rat blood.</p>	Weiss	Book	235
<p>Development  <i>The Atlas of Mouse Development</i>, 2nd ed.                      This book is the premier photomicrographic atlas for normal mouse anatomy and multiple gestational stages. Images mainly represent 2-dimensional step sections of embryos and fetuses, but some 3-dimensional plates are included to demonstrate progressive development over time of select organs.  <i>Manipulating the Mouse Embryo</i>, 3rd ed.                      This volume is an encyclopedic collection of methods used to produce genetically engineered mice, but it incorporates many standard pathology methods for morphologic analysis of mouse embryos and fetuses.</p>	Kaufman	Book	127
<p><i>Pathology of the Developing Mouse: A Systematic Approach</i>                      This book provides a methodical review of the anatomic pathology and clinical pathology techniques used for the structural analysis of mouse embryos, fetuses, and placentae. The unique feature is that patterns of common developmental defects are described and illustrated.</p>	Nagy	Book	159
<p>Pathology                      Mouse  <i>International Classification of Rodent Tumours: The Mouse</i>                      This book illustrates and describes the typical features of proliferative lesions in mice (both neoplastic and nonneoplastic). Entries include major differential diagnoses.  <i>Pathobiology of the Aging Mouse</i>                      This 2-volume set provides pathology data for many conditions of mature and senescent mice, arranged by systems. Both nonproliferative and neoplastic lesions are addressed.  <i>Pathology of Genetically Engineered Mice</i>                      This book gives an overview of the primary analytic methods used to assess genetically engineered mice for prenatal and postnatal phenotypes, with chapters describing common phenotypes for many organ systems.  <i>Pathology of Laboratory Rodents and Rabbits</i>, 3rd ed.                      This manual describes and illustrates the main lesions produced by major rodent pathogens while listing the main differential diagnoses for these diseases.  <i>Pathology of the Mouse: Reference and Atlas</i>                      This tome supplies a thorough review of the anatomy and pathology for major organ systems in the mouse. Coverage is provided for degenerative, neoplastic, and toxicant-induced lesions as well as common incidental findings.</p>	Bolon	Book	18
<p>Rat  <i>Handbook of Toxicologic Pathology</i>                      This 2-volume set contains system-specific chapters packed with normal and toxicant-altered anatomic and functional features of laboratory animals, especially rodents.  <i>Monographs on Pathology of Laboratory Animals</i>                      This multivolume collection describes system-specific pathology in rats and mice. Normal anatomy and nonproliferative and neoplastic lesions are discussed.</p>	Mohr	Book	150
	Mohr	Book <sup>a</sup>	152
	Ward	Book <sup>a</sup>	233
	Percy	Book	185
	Maronpot	Book	145
	Haschek	Book	94
	Jones	Book <sup>a</sup>	

(continued)

**Table 1.** (continued)

Title	First Author/Editor	Type	Citation
<i>Cardiovascular and Musculoskeletal Systems</i>			118
<i>Digestive System</i>			124
<i>Endocrine System</i>			115
<i>Eye and Ear</i>			119
<i>Genital System</i>			120
<i>Hematopoietic System</i>			123
<i>Integument and Mammary Glands</i>			121
<i>Nervous System</i>			122
<i>Respiratory System</i>			116
<i>Urinary System</i>			117
<i>Pathobiology of the Aging Rat</i> This 2-volume set provides pathology data for many conditions of mature and senescent rats, arranged by systems. Both nonproliferative and neoplastic lesions are addressed.	Mohr	Book <sup>a</sup>	151
<i>Pathology of the Fischer Rat: Reference and Atlas</i> This tome supplies a thorough review of the anatomy and pathology for major organ systems in the rat. Coverage is provided for degenerative, neoplastic, and toxicant-induced lesions as well as common incidental findings.	Boorman	Book <sup>a</sup>	20
<b>Organ-specific resources</b>			
<b>Nervous system</b>			
<i>The Mouse Brain in Stereotaxic Coordinates</i> , 3rd ed. This atlas shows well-annotated step sections of mouse brain features in all 3 dimensions.	Franklin	Book	74
<i>The Rat Brain in Stereotaxic Coordinates</i> , 6th ed. This atlas shows well-annotated step sections of rat brain features in all 3 dimensions. Individuals who perform rat neuropathology assessments on a regular basis may desire access to earlier editions, which present the intervening step sections.	Paxinos	Book	180
<b>Reproductive tract</b>			
"The Female Rat Reproductive Cycle: A Practical Histological Guide to Staging" This article provides a good review of the morphologic variations of different reproductive tract locations over the entire estrous cycle.	Westwood	Article	239
<b>Strain-specific physiology</b>			
"Genetic Variation Among 129 Substrains and Its Importance for Targeted Mutagenesis in Mice" This article is a powerful reminder that insufficient care in mouse husbandry can have substantial impact on genetic backgrounds and the availability of suitable control animals. The piece is important because many targeted mutations ("knockout" lines) are generated using 129 substrains.	Simpson	Article	208
"Pathology of Aging B6;129 Mice" This article presents primary lesions that frequently affect mice with this genetic background, as targeted mutations commonly introduce 129-derived embryonic stem cells into C57BL/6 blastocysts.	Haines	Article	92
"Spontaneous Lesions in Aging FVB/N Mice" This article presents primary lesions that frequently affect mice with this genetic background, as many transgenic lines are made by microinjection of engineered DNA into FVB/N oocytes.	Mahler	Article	143

<sup>a</sup>Out-of-print reference.

**Table 2.** Essential Internet Resources for Comparative Pathologists Engaged in Phenotypic Analysis of Engineered Rodents

Title	Citation
<b>Mouse</b>	
<p>"Comparative Placentation," <a href="http://placentation.ucsd.edu/">http://placentation.ucsd.edu/</a> A comprehensive site with text and images that compares placental structure among many species, including rodents. The simplest means for accessing mouse and rat data is to use the "Index" button and then scroll to the bottom to find the "Rodentia" section.</p>	15
<p>"Inbred Strains of Mice," <a href="http://www.informatics.jax.org/external/festing/mouse/STRAINS.shtml">http://www.informatics.jax.org/external/festing/mouse/STRAINS.shtml</a> This venue provides an alphabetical listing of major mouse strains with links to extensive documentation regarding biological characteristics. This site offers a thorough overview of strain-specific anatomy and physiology.</p>	67
<p>"Mouse Brain Library," <a href="http://mbl.org/">http://mbl.org/</a> This site catalogs high-resolution images and databases of brains from several mouse strains. Atlases are available with sections in 2 orientations (coronal and horizontal) for various ages.</p>	241
<p>"Mouse Genome Informatics," <a href="http://www.informatics.jax.org">http://www.informatics.jax.org</a> This portal contains genomic and phenotypic data for many mouse strains. Links provide ready access to searchable databases that match mutations (engineered and spontaneous) to phenotypes, describe gene maps and markers, and allow comparison of genomic information among species.</p>	106
<p>"Mouse Nomenclature," <a href="http://www.informatics.jax.org/mgihome/nomen/index.shtml">http://www.informatics.jax.org/mgihome/nomen/index.shtml</a> This portal provides entry to rules and guidelines for naming genes, alleles, mutations, chromosomal aberrations, and mouse and rat strains.</p>	107
<p>"Pathology of Genetically Engineered Mice," <a href="http://www.niaid.nih.gov/labsandresources/labs/aboutlabs/cmb/infectiousdiseasepathogenesissection/vetpathology/geneticallyengineeredmice/Pages/geneticallyEngineeredMice.aspx">http://www.niaid.nih.gov/labsandresources/labs/aboutlabs/cmb/infectiousdiseasepathogenesissection/vetpathology/geneticallyengineeredmice/Pages/geneticallyEngineeredMice.aspx</a> This clearinghouse provides links to numerous sites of relevance to mouse pathologists, including a link to a companion site dealing with rodent immunohistochemistry.</p>	166
<b>Pathology nomenclature</b>	
<p>These collections provide a comprehensive list of lesion names and descriptions for common abnormalities in major organs of mice and rats. The older Standardized System of Nomenclature and Diagnostic Criteria guides are currently being replaced by the INHAND documents.</p>	
<p>"Standardized System of Nomenclature and Diagnostic Criteria (SSNDC) Guides," <a href="http://www.toxpath.org/ssndc.asp">http://www.toxpath.org/ssndc.asp</a></p>	211
<p>"International Harmonization of Nomenclature and Diagnostic Criteria for Lesions in Rats and Mice," <a href="http://www.toxpath.org/inhand.asp">http://www.toxpath.org/inhand.asp</a></p>	104
<b>Phenome databases</b>	
<p>Taken together, these portals offer a comprehensive compilation of mouse physiologic data that can be searched by strain, organ system, or biological parameter. Links to protocols for acquiring such data are also given.</p>	
<p>"Europhenome," <a href="http://www.europhenome.org/databrowser/viewer.jsp">http://www.europhenome.org/databrowser/viewer.jsp</a></p>	64
<p>"Mouse Phenome Database," <a href="http://phenome.jax.org/">http://phenome.jax.org/</a></p>	108
<p>"Pathbase," <a href="http://www.pathbase.net/">http://www.pathbase.net/</a></p>	6
<p>"Phenotyping of Genetically Engineered Mice," <a href="http://dels-old.nas.edu/ilar_n/ilarjournal/47_2/html/">http://dels-old.nas.edu/ilar_n/ilarjournal/47_2/html/</a> The site provides access to the free online archive of an <i>ILAR Journal</i> issue discussing methods and policy considerations to contemplate when generating and analyzing mouse colonies with genetically engineered phenotypes.</p>	157
<p>"Revised Guides for Organ Sampling and Trimming in Rats and Mice," <a href="http://reni.item.fraunhofer.de/reni/trimming/index.php">http://reni.item.fraunhofer.de/reni/trimming/index.php</a> This site gives access to guides for organ sampling and trimming in mice and rats.</p>	158
<p>"Visible Mouse Project," <a href="http://tvmouse.compmed.ucdavis.edu/">http://tvmouse.compmed.ucdavis.edu/</a> The site is a clearinghouse for multimedia presentations describing the anatomy, physiology, histology, and pathology of the laboratory mouse, emphasizing the features of genetically engineered mice.</p>	81
<b>Rat</b>	
<p>"Rat Genome Database," <a href="http://rgd.mcw.edu/">http://rgd.mcw.edu/</a> This portal affords entry to information regarding many aspects of rat biology, including links to genomic, proteomic (pathway), functional, phenotypic, and strain data.</p>	109

by definition, it fails to provide a standard knowledge base for individuals engaged in GEM and GER phenotyping projects.

Efforts are currently underway by several organizations (eg, the Academy of Genomic Pathology, [http://ctrngenpath.net/?page\\_id=2612](http://ctrngenpath.net/?page_id=2612); the ACVP/STP Coalition for Veterinary Pathology Fellows, <http://www.vetpathcoalition.org/>) to produce more consistent training experiences in comparative pathology emphasizing rodent phenotypic analysis. However, the curricula and final product (eg, formal degree, continuing

education certificate) have yet to be defined. Therefore, individuals interested in acquiring knowledge and practical skills in comparative pathology must continue to pursue their education in a self-directed fashion for the present.

A partial remedy for the scarcity of organized training programs in rodent pathology is to devise a self-study program based on readily available resources. The current review is designed to be a multidisciplinary compilation of Internet and print resources to support a well-rounded autotutorial

**Table 3.** Age-, Disease-, and Organ-Specific References to Aid in the Phenotypic Analysis of Engineered Rodents

Subject	Citation
Anatomy	42, 45, 95, 134, 187, 207, 231
Anatomic pathology	
Macroscopic assessment	27, 66, 68
Microscopic analysis	4, 44, 87, 131, 153, 196, 209
Clinical pathology	
Baseline physiological parameters	31, 32, 137, 146, 188, 222
Sample collection	41, 100, 146
Development	
Analytical methods	10, 29, 46, 55, 159, 160
Anatomy	2, 3, 47, 49, 110, 111, 148, 172, 173, 175, 186, 198–200, 214, 221, 230, 234
Biology	52, 63, 78, 79, 85, 88, 95, 128, 129, 190, 191, 195, 197, 201, 242, 248–250
Pathology	83, 226
Disease	
Cancer	5, 17, 33, 77, 91, 103, 105, 132, 150, 154, 162, 163, 169, 203, 212, 236
Infections	8, 9, 71, 147, 213
Genetics	70, 206
Models	164
Nomenclature	139, 142
Pathology	20, 87, 89, 145, 185, 233
Phenotyping	
General	11, 12, 22, 28, 53, 102, 149, 165, 180, 217
Influence of genetic background	11, 139
Behavior	48, 155, 192
Blood	62, 65, 75, 112, 132, 146, 154, 224, 235
Cardiovascular	
Analytical methods	93, 141, 244
Anatomy	198, 232, 234, 238
Biology	114, 138
Digestive	17, 103
Endocrine	126
Eye	40, 210
Immune system	14, 38, 39, 56–61, 73, 76, 144, 154, 183, 184, 202, 218, 224, 225, 229, 240
Mammary gland	33, 34, 98
Nervous system	
Neuroanatomy	1–3, 7, 25, 50, 54, 74, 99, 133, 136, 173–181, 199, 200, 205, 219, 228, 237, 241, 246, 247
Neurochemistry	86, 111, 182, 223
Neurology	26, 50
Neurophysiology	43
Neuropathology	19, 69, 82, 84, 91, 125, 212, 236
Tissue preparation	36, 69, 97, 170
Placenta	51, 113, 161, 194
Pulmonary	141, 169
Reproductive	21, 203
Skeleton	23
Skin	156, 215
Urinary	141
Physiology	16, 37, 101, 114, 141, 189, 227
Statistics	80

curriculum in GEM and GER pathobiology and phenotyping. The cited resources are arranged in 3 tables for convenience. The first 2 tables reflect the printed materials (eg, books and journal articles; Table 1) and websites (Table 2) that we most commonly use in our daily course of comparative pathology research. Each frequently utilized reference is followed by a brief description of its content and main features. We recommend that the annotated print items in Table 1 be available in each pathologist's departmental library, and ideally in his or her office. The third table lists topics related to rodent phenotyping that occasionally require input from a comparative pathologist but fall outside the core pathology expertise expected of such individuals. These latter materials are accompanied by a list of one or more nonannotated citations that offer further discussion on such auxiliary subjects. For some topics in Table 3 (eg, neuroanatomy atlases for adult mice), multiple editions of a given book have been produced by the same authors. All editions have been included in the citation list in cases where we have found that the various versions may illuminate distinct features of the given rodent system.

A solid understanding of the foundational knowledge required for proficient GEM and GER phenotyping may be readily attained by a sincere study of the resources in the annotated lists. Sufficient time will have to be spent perusing these baseline resources so that the budding comparative pathologist becomes familiar with not only individual facts but also the location of essential topics of relevance to phenotyping GEM and GER models. Pathologists who regularly participate in rodent phenotyping will also need to engage in a lifelong quest for continuing education due to the rapid evolution of information, skills, tools, and techniques within this field.

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### References

- Allen Institute for Brain Science. *Brain Atlas Portal*. Seattle, WA: Allen Institute for Brain Science. <http://www.brain-map.org/>. Published 2009. Accessed September 21, 2009.
- Altman J, Bayer SA. *Atlas of Prenatal Rat Brain Development*. Boca Raton, FL: CRC Press; 1995.
- Alvarez-Bolado G, Swanson LW. *Developmental Brain Maps: Structure of the Embryonic Rat Brain*. New York, NY: Elsevier; 1996.
- Anonymous. *DeltaBase histology atlas*. Deltagen, Inc. website. <http://www.deltagen.com/target/histologyatlas/HistologyAtlas.html>. Published 2000–2006. Accessed January 3, 2011.
- Anonymous. *Mouse Models of Human Cancers Consortium (MMHCC) image archive*. <http://imagearchive.compmed.ucdavis.edu>. Accessed January 3, 2011.

6. Anonymous. *Pathbase: European mutant mouse pathology database*. <http://www.pathbase.net/>. Accessed January 3, 2011.
7. Ashwell KWS, Paxinos G. *Atlas of the Developing Rat Nervous System*. 3rd ed. San Diego, CA: Academic Press; 2008.
8. Baker DG. *Natural Pathogens of Laboratory Animals: Their Effects on Research*. Washington, DC: ASM Press; 2003.
9. Baker DG. Natural pathogens of laboratory mice, rats, and rabbits and their effects on research. *Clin Microbiol Rev*. 1998;**11**: 231–266.
10. Barrow MV, Taylor WJ. A rapid method for detecting malformations in rat fetuses. *J Morphol*. 1969;**127**:291–305.
11. Barthold SW. Genetically altered mice: phenotypes, no phenotypes, and faux phenotypes. *Genetica*. 2004;**122**:75–88.
12. Barthold SW. “Muromics”: genomics from the perspective of the laboratory mouse. *Comp Med*. 2002;**52**:206–223.
13. Barthold SW, Borowsky AD, Brayton C, et al. From whence will they come? A perspective on the acute shortage of pathologists in biomedical research. *J Vet Diagn Invest*. 2007;**19**:455–456.
14. Belizário JE. Immunodeficient mouse models: an overview. *Open Immunol J*. 2009;**2**:79–85.
15. Benirschke K. Comparative placentation. <http://placentation.ucsd.edu/>. Published 2007. Accessed September 1, 2007.
16. Berul CI. Electrophysiological phenotyping in genetically engineered mice. *Physiol Genomics*. 2003;**13**:207–216.
17. Boivin GP, Washington K, Yang K, et al. Pathology of mouse models of intestinal cancer: consensus report and recommendations. *Gastroenterology*. 2003;**124**:762–777.
18. Bolon B. *Pathology of the Developing Mouse: A Systematic Approach*. Boca Raton, FL: CRC Press; in press.
19. Bolon B, Garman R, Jensen K, et al. A “best practices” approach to neuropathologic assessment in developmental neurotoxicity testing—for today. *Toxicol Pathol*. 2006;**34**:296–313.
20. Boorman GA, Eustis SL, Elwell MR, et al. *Pathology of the Fischer Rat: Reference and Atlas*. San Diego, CA: Academic Press; 1990.
21. Borg CL, Wolski KM, Gibbs GM, et al. Phenotyping male infertility in the mouse: how to get the most out of a “non-performer.” *Hum Reprod Update*. 2010;**16**:205–224.
22. Boschert K. *NetVet Rodents*. <http://netvet.wustl.edu/rodents.htm>. Published 1994–1998. Accessed April 5, 2011.
23. Boskey AL, van der Meulen MCH, Wright TM. Guidelines for describing mouse skeletal phenotype. *J Orthopaed Res*. 2003;**21**:1–5.
24. Bouley D, Meyerholz D, Sellers R, et al. Where’s the mouse pathology training? *Vet Pathol*. 2009;**46**:1245–1247.
25. Brain Biodiversity Bank. *Brain Biodiversity Bank Atlases at Michigan State University*. East Lansing, MI: Michigan State University. <https://www.msu.edu/user/brains/brains/index.html>. Accessed April 5, 2011.
26. Braund KG. *Clinical Syndromes in Veterinary Neurology*. Baltimore, MD: Williams & Wilkins; 1986.
27. Brayton C, Justice M, Montgomery CA. Evaluating mutant mice: anatomic pathology. *Vet Pathol*. 2001;**38**:1–19. (Erratum in *Vet Pathol*. 2001;**38**:568)
28. Bronson RT. How to study pathologic phenotypes of knockout mice. In: Tymms MJ, Kola I, eds. *Methods in Molecular Biology: Gene Knockout Protocols*. Totowa, NJ: Humana; 2001:155–180.
29. Brown NA. Routine assessment of morphology and growth: scoring systems and measurements of size. In: Copp AJ, Cockcroft DL, eds. *Postimplantation Mammalian Embryos: A Practical Approach*. Oxford, UK: IRL Press; 1990:93–108.
30. Brown SD, Wurst W, Kühn R, et al. The functional annotation of mammalian genomes: the challenge of phenotyping. *Annu Rev Genet*. 2009;**43**:305–333.
31. Car BD, Eng VM. Special considerations in the evaluation of the hematology and hemostasis of mutant mice. *Vet Pathol*. 2001;**38**: 20–30.
32. Car BD, Eng VM, Everds NE, et al. Clinical pathology of the rat. In: Suckow MA, Weisbroth SH, Franklin CA, eds. *The Laboratory Rat*. San Diego, CA: Academic Press; 2006:127–146.
33. Cardiff RD, Anver MR, Busterson BA, et al. The mammary pathology of genetically engineered mice: the consensus report and recommendations from the Annapolis meeting. *Oncogene*. 2000;**19**:968–988.
34. Cardiff RD, Kenney N. Mouse mammary tumor biology: a short history. *Adv Cancer Res*. 2007;**98**:53–116.
35. Cardiff RD, Ward JM, Barthold SW. “One medicine—one pathology”: are veterinary and human pathology prepared? *Lab Invest*. 2008;**88**:18–26.
36. Cassella J, Hay J, Lawson S. *The Rat Nervous System: An Introduction to Preparatory Techniques*. New York, NY: John Wiley & Sons; 1997.
37. Centre for Modeling Human Diseases. Statistics for mouse physiological parameters. Toronto Centre for Phenogenomics website. [http://www.cmhd.ca/enu\\_mutagenesis/physiology\\_stats/physiology\\_stats.html](http://www.cmhd.ca/enu_mutagenesis/physiology_stats/physiology_stats.html). Published 2004. Accessed January 3, 2011.
38. Cesta MF. Normal structure, function, and histology of mucosa-associated lymphoid tissue. *Toxicol Pathol*. 2006;**34**:599–608.
39. Cesta MF. Normal structure, function, and histology of the spleen. *Toxicol Pathol*. 2006;**34**:455–465.
40. Chalupa LM, Williams RW. *Eye, Retina, and Visual System of the Mouse*. Cambridge, MA: MIT Press; 2008.
41. Chew JL, Chua KY. Collection of mouse urine for bioassays. *Lab Anim*. 2003;**32**:48–50.
42. Chiasson RB. *Laboratory Anatomy of the White Rat*. 5th ed. Boston, MA: WCB/McGraw-Hill; 1994.
43. Chudler EH. *Brain Facts and Figures*. Seattle, WA: University of Washington. <http://www.vin.com/WebLink.plx?URL=http://faculty.washington.edu/chudler/facts.html>. Published 1996. Accessed September 21, 2009.
44. Conti C, Gimenez-Conti IB, Benavides F, et al. Atlas of laboratory mouse histology. <http://ctrngenpath.net/static/atlas/mousehistology/>. Published 2004. Accessed April 1, 2011.
45. Cook MJ. *The anatomy of the laboratory mouse*. <http://www.informatics.jax.org/cookbook/>. Published 1965. Accessed January 3, 2011.
46. Copp AJ, Cockcroft DL. *Postimplantation Mammalian Embryos: A Practical Approach*. Oxford, UK: IRL Press; 1990.
47. Crawford LW, Foley JF, Elmore SA. Histology atlas of the developing mouse hepatobiliary system with emphasis on embryonic days 9.5–18.5. *Toxicol Pathol*. 2010;**38**:872–906.
48. Crawley JN. *What’s Wrong With My Mouse? Behavioral Phenotyping of Transgenic and Knockout Mice*. New York, NY: John Wiley & Sons; 2000.

49. Davidson D. *Edinburgh Mouse Atlas Project (Emap)*. Edinburgh, Scotland: United Kingdom Medical Research Council. <http://emouseatlas.org/>. Published 1994. Accessed September 21, 2009.
50. de Lahunta A, Glass E. *Veterinary Neuroanatomy and Clinical Neurology*. 3rd ed. St Louis, MO: Saunders; 2009.
51. de Rijk EPCT, van Esch E, Flik G. Pregnancy dating in the rat: placental morphology and maternal blood parameters. *Toxicol Pathol*. 2002;**30**:271–282.
52. DeSesso JM. Comparative embryology. In: Hood RD, ed. *Developmental and Reproductive Toxicology: A Practical Approach*. 2nd ed. Boca Raton, FL: CRC Press; 2006:147–197.
53. Doetschman T. Interpretation of phenotype in genetically engineered mice. *Lab Anim Sci*. 1999;**49**:137–143.
54. Dorr AE, Lerch JP, Spring S, et al. *Neuroanatomy Atlas of the C57BL/6j Mouse*. Toronto, ON: Toronto Centre for Phenogenomics. [http://www.mouseimaging.ca/research/C57Bl6j\\_mouse\\_atlas.html](http://www.mouseimaging.ca/research/C57Bl6j_mouse_atlas.html). Published 2008. Accessed September 21, 2009.
55. Downs KM, Davies T. Staging of gastrulating mouse embryos by morphological landmarks in the dissecting microscope. *Development*. 1993;**118**:1255–1266.
56. Elmore SA. Enhanced histopathology of the bone marrow. *Toxicol Pathol*. 2006;**34**:666–686.
57. Elmore SA. Enhanced histopathology of the lymph nodes. *Toxicol Pathol*. 2006;**34**:634–647.
58. Elmore SA. Enhanced histopathology of the mucosa-associated lymphoid tissue. *Toxicol Pathol*. 2006;**34**:687–696.
59. Elmore SA. Enhanced histopathology of the spleen. *Toxicol Pathol*. 2006;**34**:648–655.
60. Elmore SA. Enhanced histopathology of the thymus. *Toxicol Pathol*. 2006;**34**:656–665.
61. Elmore SA. Histopathology of the lymph nodes. *Toxicol Pathol*. 2006;**34**:425–454.
62. Emeis JJ, Jirouskova M, Muchitsch E-M, et al. A guide to murine coagulating factor structure, function, assays, and genetic alterations. *J Thromb Haemost*. 2007;**5**:670–679.
63. Erb C. Embryology and teratology. In: Suckow MA, Weisbroth SH, Franklin CA, eds. *The Laboratory Rat*. San Diego, CA: Academic Press; 2006:817–846.
64. Europhenome. *Europhenome mouse phenotyping resource*. <http://www.europhenome.org/databrowser/viewer.jsp>. Accessed January 3, 2011.
65. Everds NE. Hematology of the laboratory mouse. In: Fox JG, Barthold SW, Davisson MT, et al., eds. *The Mouse in Biomedical Research*. 2nd ed. Boston, MA: Elsevier; 2007:133–170.
66. Feldman DB, Seely JC. *Necropsy Guide: Rodents and the Rabbit*. Boca Raton, FL: CRC Press; 1988.
67. Festing MFW. *Inbred strains of mice*. Bar Harbor, ME: Mouse Genome Informatics, The Jackson Laboratory. <http://www.informatics.jax.org/external/festing/mouse/STRAINS.shtml>. Published 1998. Accessed April 5, 2011.
68. Fiette L, Slaoui M. Necropsy and sampling procedures in rodents. *Methods Mol Biol*. 2011;**691**:39–67.
69. Fix AS, Garman RH. Practical aspects of neuropathology: a technical guide for working with the nervous system. *Toxicol Pathol*. 2000;**28**:122–131.
70. Fox JG, Barthold SW, Davisson MT, et al. *The Mouse in Biomedical Research. Vol 1. History, Wild Mice, and Genetics*. 2nd ed. San Diego, CA: Academic Press; 2007.
71. Fox JG, Barthold SW, Davisson MT, et al. *The Mouse in Biomedical Research. Vol 2. Diseases*. 2nd ed. San Diego, CA: Academic Press; 2007.
72. Fox JG, Barthold SW, Davisson MT, et al. *The Mouse in Biomedical Research. Vol 3. Normative Biology, Husbandry, and Models*. 2nd ed. San Diego, CA: Academic Press; 2007.
73. Fox JG, Barthold SW, Davisson MT, et al. *The Mouse in Biomedical Research. Vol 4. Immunology*. 2nd ed. San Diego, CA: Academic Press; 2007.
74. Franklin KBJ, Paxinos G. *The Mouse Brain in Stereotaxic Coordinates*. 3rd ed. San Diego, CA: Academic Press; 2007.
75. Fredrickson TN, Harris AW. *Atlas of Mouse Hematopathology*. New York, NY: Informa Healthcare; 2000.
76. Frieke Kuper C. Histopathology of mucosa-associated lymphoid tissue. *Toxicol Pathol*. 2006;**34**:609–615.
77. Frith CH, Ward JM. *Color Atlas of Neoplastic and Non-neoplastic Lesions in Aging Mice*. Gurnee, IL: C. L. Davis, DVM Foundation; 1988.
78. Fujinaga M, Baden JM. Variation in development in rat embryos at the presomite period. *Teratology*. 1992;**45**:661–670.
79. Fujinaga M, Jackson EC, Baden JM. Interlitter variability and developmental stage of day 11 rat embryos produced by overnight and morning short-period breeding regimens. *Teratology*. 1990;**42**:535–540.
80. Gad SC. Statistics for toxicologists. In: Hayes AW, ed. *Principles and Methods of Toxicology*. 4th ed. Philadelphia, PA: Taylor & Francis; 2001:285–364.
81. Galvez J. *The visible mouse project*. <http://tvmouse.comped.ucdavis.edu/>. Accessed April 7, 2011.
82. Garman RH. Artifacts in routinely immersion fixed nervous tissue. *Toxicol Pathol*. 1990;**18**:149–153.
83. German Federal Institute for Risk Assessment. *DevTox*. <http://www.devtox.org/>. Published 2009. Accessed January 3, 2011.
84. Goelz MF, Mahler J, Harry J, et al. Neuropathologic findings associated with seizures in FVB mice. *Lab Anim Sci*. 1998;**48**:34–37.
85. Goldman AS. Critical periods of prenatal toxicity. *Clin Perinatol*. 1979;**6**:203–218.
86. Goldowitz D. Histological phenotyping of the mouse brain. Tennessee Mouse Genome Consortium. <http://neurophenotyping.uthsc.edu/index%20.html>. Published 2005. Accessed January 3, 2011.
87. Gopinath C, Prentice DE, Lewis DJ. *Atlas of Experimental Toxicological Pathology*. Lancaster, UK: MTP Press; 1987.
88. Gossler A. Early mouse development. In: Hennig W, ed. *Early Embryonic Development of Animals*. Berlin, Germany: Springer-Verlag; 1992:151–201.
89. Greaves P. *Histopathology of Preclinical Toxicity Studies: Interpretation and Relevance in Drug Safety Evaluation*. 3rd ed. New York, NY: Academic Press; 2007.
90. Greene EC. *Anatomy of the Rat*. Braintree, MA: Braintree Scientific, Inc.; 1935.
91. Gutmann DH, Baker S, Giovannini M, et al. Mouse models of human cancer consortium symposium on nervous system tumors. *Cancer Res*. 2003;**63**:3001–3004.



92. Haines DC, Chattopadhyay S, Ward JM. Pathology of aging B6; 129 mice. *Toxicol Pathol.* 2001;**29**:653–661.
93. Hartley CJ, Taffet GE, Reddy AK, et al. Noninvasive cardiovascular phenotyping in mice. *ILAR J.* 2002;**43**:147–158.
94. Haschek WM, Rousseaux CG, Wallig MA. *Handbook of Toxicologic Pathology.* 2nd ed. San Diego, CA: Academic Press; 2002.
95. Hebel R, Stromberg MW. *Anatomy and Embryology of the Rat.* Wörthsee, Germany: BioMed Verlag; 1986.
96. Hedrich H. *The Laboratory Mouse.* San Diego, CA: Academic Press; 2004.
97. Heffner TG, Hartman JA, Seiden LS. A rapid method for the regional dissection of the rat brain. *Pharmacol Biochem Behav.* 1980;**13**:453–456.
98. Hennighausen L, Wagner U. *Biology of the mammary gland.* <http://mammary.nih.gov/index.html>. Accessed April 1, 2011.
99. Hof PR, Young PR, Bloom FE, et al. *Comparative Cytoarchitectonic Atlas of the C57BL/6 and 129/Sv Mouse Brains.* Philadelphia, PA: Elsevier; 2000.
100. Hoff J. Methods of blood collection in the mouse. *Lab Anim.* 2000;**29**:47–53.
101. Hoyt RFJ, Hawkins JV, St Claire MB, Kennett MJ. Mouse physiology. In: Fox JG, Barthold SW, Davisson MT, et al., eds. *The Mouse in Biomedical Research.* 2nd ed. Boston, MA: Elsevier; 2007: 23–90.
102. Hrabé de Angelis M, Chambon P, Brown S. *Standards of Mouse Model Phenotyping.* Weinheim, Germany: Wiley-VCH Verlag GmbH & Co.; 2006.
103. Hruban RH, Adsay NV, Albores-Saavedra J, et al. Pathology of genetically engineered mouse models of pancreatic exocrine cancer: consensus report and recommendations. *Cancer Res.* 2006;**66**:95–106.
104. INHAND. *International harmonization of nomenclature and diagnostic criteria for lesions in rats and mice.* <http://www.tox-path.org/inhand.asp>. Accessed April 5, 2011.
105. Ito N, Hayashi Y. Electronic color atlas: histopathology of chemically induced tumors. <http://ctrngenpath.net/static/atlas/chemicallyinducedtumors/files/index.html>. 1987.
106. Jackson Laboratory. *Mouse genome informatics.* <http://www.informatics.jax.org>. Published 2010. Accessed January 3, 2011.
107. Jackson Laboratory. *Mouse nomenclature.* <http://www.informatics.jax.org/mgihome/nomen/index.shtml>. Published 2010. Accessed January 3, 2011.
108. Jackson Laboratory. *Mouse phenome database.* <http://phenome.jax.org/>. Published 2001–2010. Accessed January 3, 2011.
109. Jacob HJ. Rat genome database. Medical College of Wisconsin website. <http://rgd.mcw.edu/>. Accessed January 3, 2011.
110. Jacobson AG, Tam PPL. Cephalic neurulation in the mouse embryo analyzed by SEM and morphometry. *Anat Rec.* 1982;**203**:375–396.
111. Jacobowitz DM, Abbott LC. *Chemoarchitectonic Atlas of the Developing Mouse Brain.* Boca Raton, FL: CRC Press; 1998.
112. Jirouskova M, Shet AS, Johnson GJ. A guide to murine platelet structure, function, assays, and genetic alterations. *J Thromb Haemost.* 2007;**5**:661–669.
113. Jollie WP. Development, morphology, and function of the yolk-sac placenta of laboratory rodents. *Teratology.* 1990;**41**:361–381.
114. Jones GLAH, Sang E, Goddard C, et al. A functional analysis of mouse models of cardiac disease through metabolic profiling. *J Biol Chem.* 2005;**280**:7530–7539.
115. Jones TC, Capen CC, Mohr U. *Monographs on Pathology of Laboratory Animals: Endocrine System.* 2nd ed. Springer-Verlag, Berlin; 1996.
116. Jones TC, Dungworth DL, Mohr U. *Monographs on Pathology of Laboratory Animals: Respiratory System.* 2nd ed. Springer-Verlag, Berlin; 1996.
117. Jones TC, Hard GC, Mohr U. *Monographs on Pathology of Laboratory Animals: Urinary System.* 2nd ed. Springer-Verlag, Berlin; 1998.
118. Jones TC, Mohr U, Hunt RD. *Monographs on Pathology of Laboratory Animals: Cardiovascular and Musculoskeletal Systems.* Springer-Verlag, Berlin; 1991.
119. Jones TC, Mohr U, Hunt RD. *Monographs on Pathology of Laboratory Animals: Eye and Ear.* Springer-Verlag, Berlin; 1991.
120. Jones TC, Mohr U, Hunt RD. *Monographs on Pathology of Laboratory Animals: Genital System.* Springer-Verlag, Berlin; 1987.
121. Jones TC, Mohr U, Hunt RD. *Monographs on Pathology of Laboratory Animals: Integument and Mammary Glands.* Springer-Verlag, Berlin; 1989.
122. Jones TC, Mohr U, Hunt RD. *Monographs on Pathology of Laboratory Animals: Nervous System.* Springer-Verlag, Berlin; 1988.
123. Jones TC, Mohr U, Hunt RD, et al. *Monographs on Pathology of Laboratory Animals: Hematopoietic System.* Springer-Verlag, Berlin; 1990.
124. Jones TC, Popp JA, Mohr U. *Monographs on Pathology of Laboratory Animals: Digestive System.* 2nd ed. Springer-Verlag, Berlin; 1997.
125. Jortner BS. The return of the dark neuron: a histological artifact complicating contemporary neurotoxicologic evaluation. *Neurotoxicology.* 2006;**27**:628–634. (Comment in *Neurotoxicology.* 2006;**27**:1126)
126. Kaufman M, Nikitin AY, Sundberg JP. *Histologic Basis of Mouse Endocrine System Development: A Comparative Analysis.* Boca Raton, FL: CRC Press; 2010.
127. Kaufman MH. *The Atlas of Mouse Development.* 2nd ed. San Diego, CA: Academic Press; 1992.
128. Kaufman MH. Mouse and human embryonic development: a comparative overview. In: Strachan T, Lindsay S, Wilson DI, eds. *Molecular Genetics of Early Human Development.* Oxford, UK: Bios Scientific; 1997:77–110.
129. Kaufman MH, Bard JBL. *The Anatomical Basis of Mouse Development.* San Diego, CA: Academic Press; 1999.
130. Kim IY, Shin JH, Seong JK. Mouse phenogenomics, toolbox for functional annotation of human genome. *BMB Rep.* 2010;**43**: 79–90.
131. Kittel B, Ruehl-Fehlert C, Morawietz G, et al. Revised guides for organ sampling and trimming in rats and mice: part 2. *Exp Toxicol Pathol.* 2004;**55**:413–431.
132. Kogan SC, Ward JM, Anver MR, et al. Hematopathology Subcommittee of the Mouse Models of Human Cancers Consortium.

- Bethesda proposals for classification of nonlymphoid hematopoietic neoplasms in mice. *Blood*. 2002;**100**:238–245.
133. Kolb B, Sutherland RJ, Nonneman AJ, Whishaw IQ. Asymmetry in the cerebral hemispheres of the rat, mouse, rabbit, and cat: the right hemisphere is larger. *Exp Neurol*. 1982;**78**:348–359.
  134. Komárek V. Gross anatomy. In: Fox JG, Barthold SW, Davisson MT, et al., eds. *The Mouse in Biomedical Research*. 2nd ed. Boston, MA: Elsevier; 2007:1–22.
  135. Krinke G. *The Laboratory Rat*. San Diego, CA: Academic Press; 2000.
  136. Kruger L, Saporta S, Swanson LW. *Photographic Atlas of the Rat Brain*. Cambridge, MA: Cambridge University Press; 1995.
  137. LaBorde JB, Wall KS, Bolon B, et al. Haematology and serum chemistry parameters of the pregnant rat. *Lab Anim*. 1999;**33**: 275–287.
  138. Leu M, Ehler E, Perriard JC. Characterisation of postnatal growth of the murine heart. *Anat Embryol (Berl)*. 2001;**204**: 217–224.
  139. Linder CC, Davisson MT. Strains, stocks, and mutant mice. In: Hedrich H, Bullock G, eds. *The Laboratory Mouse*. San Diego, CA: Academic Press; 2004: 25–46.
  140. Loeb WF, Quimby FW. *The Clinical Chemistry of Laboratory Animals*. 2nd ed. Philadelphia, PA: Taylor & Francis; 1999.
  141. Lorenz JN. A practical guide to evaluating cardiovascular, renal, and pulmonary function in mice. *Am J Physiol Regul Integr Comp Physiol*. 2002;**282**:R1565–R1582.
  142. Lyon MF, Rastan S, Brown SDM. *Genetic Variants and Strains of the Laboratory Mouse*. 3rd ed. Oxford, UK: Oxford University Press; 1996.
  143. Mahler JF, Stokes W, Mann PC, et al. Spontaneous lesions in aging FVB/N mice. *Toxicol Pathol*. 1996;**24**:710–716.
  144. Maronpot RR. Enhanced histopathology of lymphoid tissues. *Toxicol Pathol*. 2006;**34**:631–633.
  145. Maronpot RR. *Pathology of the Mouse: Reference and Atlas*. Vienna, IL: Cache River Press; 1999.
  146. McGarry MP, Protheroe CA, Lee JJ. *Mouse Hematology: A Laboratory Manual*. Woodbury, NY: Cold Spring Harbor Laboratory Press; 2010.
  147. Mendlowitz L. *Diseases of Research Animals*. Columbia, MO: University of Missouri, Research Animal Diagnostic Laboratory. <http://www.radil.missouri.edu/info/dora/Dora.htm>. Published 2002. Accessed January 3, 2011.
  148. Menegola E, Broccia ML, Giavini E. Atlas of rat fetal skeleton double stained for bone and cartilage. *Teratology*. 2001;**64**: 125–133.
  149. Mercer EH. The whole mouse catalog. <http://wmc.rodentia.com/>. Published 1995. Accessed September 21, 2009.
  150. Mohr U. *International Classification of Rodent Tumours: The Mouse*. Berlin, Germany: Springer-Verlag; 2001.
  151. Mohr U, Dungworth DL, Capen CC. *Pathobiology of the Aging Rat*. Washington, DC: ILSI Press; 1992.
  152. Mohr U, Dungworth DL, Capen CC, et al. *Pathobiology of the Aging Mouse*. Washington, DC: ILSI Press; 1996.
  153. Morawietz G, Ruehl-Fehlert C, Kittel B, et al. Revised guides for organ sampling and trimming in rats and mice: part 3. *Exp Toxicol Pathol*. 2004;**55**:433–449.
  154. Morse HC 3rd, Anver MR, Fredrickson TN, et al. Hematopathology Subcommittee of the Mouse Models of Human Cancers Consortium. Bethesda proposal for classification of lymphoid neoplasms in mice. *Blood*. 2002;**100**:246–258.
  155. Moser VC. The functional observational battery in adult and developing rats. *Neurotoxicology*. 2000;**21**:989–996.
  156. Muller-Rover S, Handjiski B, van der Veen C, et al. A comprehensive guide for the accurate classification of murine hair follicles in distinct hair cycle stages. *J Invest Dermatol*. 2001;**117**:3–15.
  157. [Multiple authors]. Phenotyping of genetically engineered mice [ILAR J, vol 47, no. 2]. Institute for Laboratory Animal Research website. [http://dels-old.nas.edu/ilar\\_n/ilarjournal/47\\_2/html/](http://dels-old.nas.edu/ilar_n/ilarjournal/47_2/html/). Published 2006. Accessed January 3, 2011.
  158. [Multiple authors]. Revised guides for organ sampling and trimming in rats and mice. Registry Nomenclature Information System website. <http://reni.item.fraunhofer.de/reni/trimming/index.php>. Published 2003–2004. Accessed April 5, 2011, 2011.
  159. Nagy A, Gertsenstein M, Vintersten K, Behringer R. *Manipulating the Mouse Embryo*. 3rd ed. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 2003.
  160. Narotsky MG, Rogers JM. Examination of the axial skeleton of fetal rodents. *Methods Mol Biol*. 2000;**135**:139–150.
  161. Natale DRC, Starovic M, Cross JC. Phenotypic analysis of the mouse placenta. In: Soares MJ, Hunt JS, eds. *Placenta and Trophoblast: Methods and Protocols*. Totowa, NJ: Humana Press; 2006:275–293.
  162. National Cancer Institute. Cancer images database. <http://cancerimages.nci.nih.gov/caIMAGE/>. Accessed April 5, 2011.
  163. National Cancer Institute. Cancer models database. <http://cancermodels.nci.nih.gov/camod/>. Accessed April 5, 2011.
  164. National Cancer Institute. eMICE: electronic models information, communication, and education. <http://emice.nci.nih.gov/>. Accessed April 5, 2011.
  165. National Institute of Allergy and Infectious Diseases, Comparative Medicine Branch, Veterinary Pathology Division. Pathology and biology of genetically engineered mice. <http://www.niaid.nih.gov/labsandresources/labs/aboutlabs/cmb/infectious-diseasepathogenesissection/vetpathology/geneticallyengineeredmice/Pages/geneticallyEngineeredMice.aspx>. Published 2007. Accessed April 5, 2011.
  166. National Institute of Allergy and Infectious Diseases, Comparative Medicine Branch, Veterinary Pathology Division. Pathology of genetically engineered mice. <http://www.niaid.nih.gov/labsandresources/labs/aboutlabs/cmb/infectiousdiseasepathogenesissection/vetpathology/geneticallyengineeredmice/Pages/geneticallyEngineeredMice.aspx>. Published 2011. Accessed April 5, 2011.
  167. Nebert DW, Dalton TP, Stuart GW, et al. “Gene-swap knock-in” cassette in mice to study allelic differences in human genes. *Ann N Y Acad Sci*. 2000;**919**:148–170.
  168. Nebert DW, Duffy JJ. How knockout mouse lines will be used to study the role of drug-metabolizing enzymes and their receptors during reproduction and development, and in environmental toxicity, cancer, and oxidative stress. *Biochem Pharmacol*. 1997;**53**:249–254.

169. Nikitin AY, Alcaraz A, Anver MR, et al. Classification of proliferative pulmonary lesions of the mouse: recommendations of the Mouse Models of Human Cancers Consortium. *Cancer Res.* 2004;**64**:2307–2316.
170. Palmblad K, Erlandsson-Harris H, Tracey KJ, et al. Dynamics of early synovial cytokine expression in rodent collagen-induced arthritis: a therapeutic study using a macrophage-deactivating compound. *Am J Pathol.* 2001;**158**:491–500.
171. Papaioannou VE, Behringer RR. *Mouse Phenotypes: A Handbook of Mutation Analysis.* Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press; 2005.
172. Paxinos G. *The Rat Nervous System.* 3rd ed. San Diego, CA: Academic Press; 2004.
173. Paxinos G, Ashwell KWS, Törk I. *Atlas of the Developing Rat Nervous System.* 2nd ed. San Diego, CA: Academic Press; 1994.
174. Paxinos G, Franklin KBJ. *The Mouse Brain in Stereotaxic Coordinates.* 2nd ed. New York, NY: Elsevier; 2001.
175. Paxinos G, Halliday G, Watson C, et al. *Atlas of the Developing Mouse Brain at E17.5, P0, and P6.* San Diego, CA: Academic Press; 2007.
176. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* Sydney, Australia: Academic Press; 1982.
177. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* 2nd ed. San Diego, CA: Academic Press; 1986.
178. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* 3rd ed. San Diego, CA: Academic Press; 1997.
179. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* 4th ed. San Diego, CA: Academic Press; 1998.
180. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* 5th ed. San Diego, CA: Academic Press; 2004.
181. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates.* 6th ed. San Diego, CA: Academic Press; 2007.
182. Paxinos G, Watson C, Carrive P, et al. *Chemoarchitectonic Atlas of the Rat Brain.* 2nd ed. San Diego, CA: Academic Press; 2008.
183. Pearse G. Histopathology of the thymus. *Toxicol Pathol.* 2006;**34**:515–547.
184. Pearse G. Normal structure, function and histology of the thymus. *Toxicol Pathol.* 2006;**34**:504–514.
185. Percy DH, Barthold SW. *Pathology of Laboratory Rodents and Rabbits.* 3rd ed. Ames, IA: Iowa State University Press; 2007.
186. Petiet AE, Kaufman MH, Goddeeris MM, et al. High-resolution magnetic resonance histology of the embryonic and neonatal mouse: a 4D atlas and morphologic database. *Proc Natl Acad Sci U S A.* 2008;**105**:12331–12336.
187. Popesko P, Rajtova V, Horak J. *Colour Atlas of Anatomy of Small Laboratory Animals.* London, UK: WB Saunders; 2003.
188. Quimby FW, Luong RH. Clinical chemistry of the laboratory mouse. In: Fox JG, Barthold SW, Davisson MT, et al., eds. *The Mouse in Biomedical Research.* 2nd ed. Boston, MA: Elsevier; 2007:171–216.
189. Rao S, Verkman AS. Analysis of organ physiology in transgenic mice. *Am J Physiol Cell Physiol.* 2000;**279**:C1–C18.
190. Rice D, Barone SJ. Critical periods of vulnerability for the developing nervous system: evidence from humans and animal models. *Environ Health Perspect.* 2000;**108**(suppl 3): 511–533.
191. Rodier PM. Chronology of neuron development: animal studies and their clinical implications. *Dev Med Child Neurol.* 1980;**22**: 525–545.
192. Rogers DC, Fisher EM, Brown SD, et al. Behavioral and functional analysis of mouse phenotype: SHIRPA, a proposed protocol for comprehensive phenotype assessment. *Mamm Genome.* 1997;**8**:711–713.
193. Rosenthal N, Brown S. The mouse ascending: perspectives for human-disease models. *Nat Cell Biol.* 2007;**9**:993–999.
194. Rossant J, Cross JC. Placental development: lessons from mouse mutants. *Nat Rev Genet.* 2001;**2**:538–548.
195. Rossant J, Tam PPL. *Mouse Development: Patterning, Morphogenesis, Organogenesis.* San Diego, CA: Academic Press; 2002.
196. Ruehl-Fehlert C, Kittel B, Morawietz G, et al. Revised guides for organ sampling and trimming in rats and mice: part 1. *Exp Toxicol Pathol.* 2003;**55**:91–106.
197. Rugh R. *The Mouse: Its Reproduction and Development.* Oxford, UK: Oxford University Press; 1990.
198. Savolainen SM, Foley JF, Elmore SA. Histology atlas of the developing mouse heart with emphasis on E11.5 to E18.5. *Toxicol Pathol.* 2009;**37**:395–414.
199. Schambra U. *Prenatal Mouse Brain Atlas.* New York, NY: Springer; 2008.
200. Schambra UB. Electronic prenatal mouse brain atlas. <http://www.epmba.org/>. Published 2007. Accessed September 21, 2009.
201. Schneider BF, Norton S. Equivalent ages in rat, mouse and chick embryos. *Teratology.* 1979;**19**:273–278.
202. Seymour R, Sundberg JP, Hogenesch H. Abnormal lymphoid organ development in immunodeficient mutant mice. *Vet Pathol.* 2006;**43**:401–423.
203. Shappell SB, Thomas GV, Roberts RL, et al. Prostate pathology of genetically engineered mice: definitions and classification. The consensus report from the Bar Harbor meeting of the Mouse Models of Human Cancer Consortium Prostate Pathology Committee. *Cancer Res.* 2004;**64**:2270–2305.
204. Sharp JJ, Mobraaten LE, Bedigian HG. Mutant mouse resources. In: Ward JM, Mahler JF, Maronpot RR, et al., eds. *Pathology of Genetically Engineered Mice.* Ames, IA: Iowa State University Press; 2000:3–9.
205. Sidman RL, Kosaras B, Misra B, et al. *High resolution mouse brain atlas.* <http://www.hms.harvard.edu/research/brain/atlas.html#>. Accessed January 3, 2011.
206. Silver LM. *Mouse Genetics: Concepts and Applications.* Oxford, UK: Oxford University Press; 2008.
207. Silverman S, Tell LA. *Radiology of Rodents, Rabbits, and Ferrets: An Atlas of Normal Anatomy and Positioning.* Philadelphia, PA: Elsevier; 2005.
208. Simpson EM, Linder CC, Sargent EE, et al. Genetic variation among 129 substrains and its importance for targeted mutagenesis in mice. *Nat Genet.* 1997;**16**:19–27.
209. Slaoui M, Fiette L. Histopathology procedures: from tissue sampling to histopathological evaluation. *Methods Mol Biol.* 2011;**691**:69–82.
210. Smith RS. *Systematic Evaluation of the Mouse Eye: Anatomy, Pathology, and Biomethods.* Boca Raton, FL: CRC Press; 2002.

211. Society of Toxicologic Pathology. Standardized system of nomenclature and diagnostic criteria (SSNDC) guides. <http://www.toxpath.org/ssndc.asp>. Accessed April 5, 2011.
212. Stemmer-Rachamimov AO, Louis DN, Nielsen GP, et al. Comparative pathology of nerve sheath tumors in mouse models and humans. *Cancer Res.* 2004;**64**:3718–3724.
213. Suckow MA, Weisbroth SH, Franklin CA. *The Laboratory Rat*. San Diego, CA: Academic Press; 2006.
214. Sulik KK, Bream PRJ. *Embryo Images: Normal and Abnormal Mammalian Development*. Chapel Hill, NC: University of North Carolina. [http://www.med.unc.edu/embryo\\_images/](http://www.med.unc.edu/embryo_images/). Accessed September 21, 2009.
215. Sundberg JP. *Handbook of Mouse Mutations With Skin and Hair Abnormalities: Animal Models and Biomedical Tools*. Boca Raton, FL: CRC Press; 1994.
216. Sundberg JP, Boggess D. *Systematic Approach to Evaluation of Mouse Mutations*. Boca Raton, FL: CRC Press; 2000.
217. Sundberg JP, Ichiki T. *Genetically Engineered Mice Handbook*. Boca Raton, FL: CRC Press; 2006.
218. Suttie AW. Histopathology of the spleen. *Toxicol Pathol.* 2006;**34**:466–503.
219. Swanson LW. *Brain Maps: Structure of the Rat Brain*. 3rd ed. San Diego, CA: Academic Press; 2003.
220. Tang T, Li L, Tang J, et al. A mouse knockout library for secreted and transmembrane proteins. *Nat Biotechnol.* 2010;**28**:749–755.
221. Theiler K. *The House Mouse: Atlas of Embryonic Development*. New York, NY: Springer-Verlag; 1989.
222. Thrall MA, Baker DC, Campbell TW, et al. *Veterinary Hematology and Clinical Chemistry*. Baltimore, MD: Lippincott Williams & Wilkins; 2004.
223. Tohyama M, Takatsuji K. *Atlas of Neuroactive Substances and Their Receptors in the Rat*. Oxford, UK: Oxford University Press; 1993.
224. Travlos GS. Histopathology of bone marrow. *Toxicol Pathol.* 2006;**34**:566–598.
225. Travlos GS. Normal structure, function, and histology of the bone marrow. *Toxicol Pathol.* 2006;**34**:548–565.
226. Turgeon B, Meloche S. Interpreting neonatal lethal phenotypes in mouse mutants: insights into gene function and human diseases. *Physiol Rev.* 2009;**89**:1–26.
227. Ullman-Culleré MH, Foltz CJ. Body condition scoring: a rapid and accurate method for assessing health status in mice. *Lab Anim Sci.* 1999;**49**:319–323.
228. Valverde F. *Golgi Atlas of the Postnatal Mouse Brain*. New York, NY: Springer; 2004.
229. Van den Broeck W, Derore A, Simoens P. Anatomy and nomenclature of murine lymph nodes: descriptive study and nomenclature standardization in BALB/cAnNCrI mice. *J Immunol Methods.* 2006;**312**:12–19.
230. Wainwright P, Deeks S. A comparison of corpus callosum development in the BALB/cCF and C57BL/6J inbred mouse strains. *Growth.* 1984;**48**:192–197.
231. Walker WFJ, Homberger DG. *Anatomy and Dissection of the Rat*. New York, NY: WH Freeman & Co; 1998.
232. Waller BRR, Wessels A. Cardiac morphogenesis and dysmorphogenesis: an immunohistochemical approach. *Methods Mol Biol.* 2000;**135**:151–161.
233. Ward JM, Mahler JF, Maronpot RR, Sundberg JP. *Pathology of Genetically Engineered Mice*. Ames, IA: Iowa State University Press; 2000.
234. Webb S, Brown NA, Anderson RH. The structure of the mouse heart in late fetal stages. *Anat Embryol (Berl)* 1996;**194**:37–47.
235. Weiss DJ, Wardrop KJ. *Schalm's Veterinary Hematology*. 6th ed. New York, NY: Wiley-Blackwell; 2010.
236. Weiss WA, Israel M, Cobbs C, et al. Neuropathology of genetically engineered mice: consensus report and recommendations from an international forum. *Oncogene.* 2002;**21**:7453–7463.
237. Welker W, Johnson JI, Noe A. Comparative mammalian brain collections. <http://www.brainmuseum.org/>. Accessed September 21, 2009.
238. Wessels A, Markwald R. Cardiac morphogenesis and dysmorphogenesis: I. Normal development. *Methods Mol Biol.* 2000;**136**:239–259.
239. Westwood FR. The female rat reproductive cycle: a practical histological guide to staging. *Toxicol Pathol.* 2008;**36**:375–384.
240. Willard-Mack CL. Normal structure, function, and histology of lymph nodes. *Toxicol Pathol.* 2006;**34**:409–424.
241. Williams RW, Rosen GD. *Mouse Brain Library*. <http://www.mbl.org/>. Published 1999. Accessed January 10, 2011.
242. Wood SL, Beyer BK, Cappon GD. Species comparison of postnatal CNS development: functional measures. *Birth Defects Res B Dev Reprod Toxicol.* 2003;**68**:391–407.
243. Xie W, Barwick JL, Downes M, et al. Humanized xenobiotic response in mice expressing nuclear receptor SXR. *Nature.* 2000;**406**:435–439.
244. Xu Q. *A Handbook of Mouse Models of Cardiovascular Disease*. West Sussex, England: John Wiley & Sons Ltd.; 2006.
245. Zambrowicz BP, Sands AT. Knockouts model the 100 best-selling drugs: will they model the next 100? *Nat Rev Drug Discov.* 2003;**2**:38–51.
246. Zeman W, Innes JRM. *Craige's Neuroanatomy of the Rat*. New York, NY: Academic Press; 1963.
247. Zilles K. *The Cortex of the Rat: A Stereotaxic Atlas*. New York, NY: Springer-Verlag; 1985.
248. Zoetis T, Hurtt ME. Species comparison of anatomical and functional renal development. *Birth Defects Res B Dev Reprod Toxicol.* 2003;**68**:111–120.
249. Zoetis T, Hurtt ME. Species comparison of lung development. *Birth Defects Res B Dev Reprod Toxicol.* 2003;**68**:121–124.
250. Zoetis T, Tassinari MS, Bagi C, et al. Species comparison of postnatal bone growth and development. *Birth Defects Res B Dev Reprod Toxicol.* 2003;**68**:86–110.